

OPTIMIZATION OF EXTRACTION PARAMETERS OF SELECTED
MALAYSIAN PLANTS TOWARDS ANTIUROLITHIATIC ACTIVITIES (*IN-
VITRO*)

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I hereby declare that the work in this Master's Thesis is my own except the quotations and summaries which have been duly acknowledged.

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ABSTRACT

Urolithiasis has become a worldwide problem and its management depends on surgical procedures that are costly and cause reoccurrence. *Anacardium occidentale* (gajus), *Ananas comosus* (nanas), *Aquilaria malaccensis* (karas), *Centella asiatica* (pegaga), *Ceiba petandra* (kekabu), *Euphorbia hirta* (ara tanah), *Ficus carica* (ara), *Melastoma malabthricum* (senduduk), *Piper sarmentosum* (kaduk) and *Tradescantia zebrina* (kura-kura air) have been used traditionally to treat urolithiasis. However, no scientific data has been recorded on the extraction parameters. Thus, the objective of this study is to assess for the best ethnobotanical plant extract with good antiurolithiatic properties, followed by optimization of extraction parameters of selected plant extracts. The antiurolithiatic activities conducted were turbidity and titrimetric assays (*in-vitro*). Ethnobotanical plant extracts assessment revealed *A. occidentale* and *A. malaccensis* had the best antiurolithiatic activities as both plants had no significant difference with standard drugs ($p>0.05$). The extraction parameters studied were solvent concentration, extraction temperature and time that was screened using two-level factorial design and optimized by response surface methodology and central composite design. The results showed that optimum extraction condition for *A. occidentale* extract (0.4% ethanol, 31.5°C, 30 minutes) on both antiurolithiatic activities were exhibited $85.57\pm0.43\%$ (turbidity) and $96.48\pm0.70\%$ (titrimetric). Meanwhile, optimum condition of *A. malaccensis* extract (100% ethanol, 30.0°C, 30 minutes) demonstrated $83.58\pm0.75\%$ (turbidity) and $86.57\pm0.80\%$ (titrimetric). Phenols, alkaloids, saponins, flavonoids, tannins and terpenoids were identified in both optimized extracts and all have positive correlation on both antiurolithiatic assays except flavonoids. Toxicity testing using brine shrimp lethality assay presented non-toxic effect on optimized *A. occidentale* (LC_{50} , 1412.50 μ g/mL) but *A. malaccensis* (LC_{50} , 30.50 μ g/mL) revealed toxic effects. This study has given basic scientific evidence that optimum extraction condition is necessary to obtain optimum antiurolithiatic activity.

ABSTRAK

Urolithiasis telah menjadi permasalahan di seluruh dunia dan pengurusannya bergantung pada prosedur pembedahan yang mahal dan sering menyebabkan pembentukan semula. *Anacardium occidentale* (gajus), *Ananas comosus* (nanas), *Aquilaria malaccensis* (karas), *Centella asiatica* (pegaga), *Ceiba petandra* (kekabu), *Euphorbia hirta* (ara tanah), *Ficus carica* (ara), *Melastoma malabthricum* (senduduk), *Piper sarmentosum* (kaduk) dan *Tradescantia zebrina* (kura-kura air) telah digunakan secara tradisional untuk merawat urolithiatik. Bagaimanapun, tiada data saintifik yang direkodkan mengenai parameter pengekstrakan. Oleh itu, objektif kajian ini adalah untuk menyaring ekstrak herba etnobotanik terbaik dengan sifat antiurolithiatik yang baik, diikuti dengan pengoptimuman parameter pengekstrakan terhadap ekstrak terpilih. Kaedah antiurolithiatik yang dilakukan adalah kekeruhan dan titrimetrik (*in-vitro*). Analisis awal memaparkan aktiviti antiurolithiatik terbaik pada *A. occidentale* dan *A. malaccensis* kerana tiada perbezaan yang ketara dengan ubat standard, ($p > 0.05$). Parameter pengekstrakan yang dikaji adalah kepekatan pelarut, suhu dan masa pengekstrakan yang disaring melalui kaedah faktorial dua peringkat dan dioptimumkan dengan kaedah permukaan tindak balas dan komposit pusat. Hasil kajian mendapati bahawa keadaan pengekstrakan optimum untuk aktiviti antiurolithiatik bagi ekstrak *A. occidentale* (0.4% etanol, 31.5°C, 30 minit) memperoleh 85.57±0.43% (kekeruhan) dan 96.48±0.70% (titrimetrik). Sementara itu, keadaan optimum ekstrak *A. malaccensis* (100% etanol, 30°C, 30 minit) menunjukkan 83.58±0.75% (kekeruhan) dan 86.57±0.80% (titrimetrik). Fenol, alkaloid, saponin, flavonoid, tanin dan terpenoid dikenal pasti dalam kedua-dua ekstrak yang dioptimumkan dan kesemuanya mempunyai hubungan positif dengan ujian antiurolithiatik kecuali flavonoid. Ujian ketoksikan mendapati kesan tidak toksik pada *A. occidentale* yang dioptimumkan (LC₅₀, 1412.50µg/mL) tetapi *A. malaccensis* (LC₅₀, 30.50µg/mL) menunjukkan kesan toksik. Kajian ini telah memberikan bukti saintifik asas bahawa keadaan pengekstrakan optimum diperlukan untuk mendapatkan aktiviti antiurolithiatik yang optimum.

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LIST OF SYMBOLS AND ABBREVIATIONS

%	-	Percent
°C	-	Degree celcius
cm	-	Centimeter
g	-	Gram
M	-	Molarity
mg	-	Milligram
mL	-	Milliliter
mm	-	Millimeter
N	-	Normality
µg	-	Microgram
µL	-	Microliter
ANOVA	-	Analysis of variance
BSLA	-	Brine shrimp lethality assay
CaOx	-	Calcium oxalate
CCD	-	Central composite design
CV	-	Coefficient of variation
ESWL	-	Extracorporeal shock wave lithotripsy
GAE	-	Gallic acid equivalent
LC	-	Lethality concentration
PNL	-	Percutaneous nephrolithotomy
RE	-	Rutin equivalent
RSM	-	Response surface methodology
TAE	-	Tannic acid equivalent
TFC	-	Total flavonoid content
TPC	-	Total phenolic content

- URS - Uretoscopy
- WHO - World Health Organization



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CHAPTER 1

INTRODUCTION

1.1 Research background

In this century, medicinal plant therapies that have been acknowledged by traditional practitioners to cure various diseases has been well adopted and adapted by today communities all over the world. Globally, herbal medicines are growing fast nowadays might be because they are cost effective, eco-friendly, readily available and culturally acceptable as well as high margin of safety with minimal side effects as compared to synthetic drugs (Arya, Pandey, & Verma, 2017; Biglarkhani *et al.*, 2017; Jain, 2003). Besides that, medicinal plants have a rich source of phytochemicals and allopathic constituents that have been conventionally used for treating ailments including urolithiasis which is characterized by the formation of stones in the urinary system (Ahmed, Hasan & Mahmood, 2016; Yadav *et al.*, 2011; Ahmad & Ismail, 2003).

Malaysia is well-known to have wide variety of medicinal plant sources that may mediate urolithiasis. Diverse ethnicity in Malaysian communities such as Malay, Chinese, Indian and indigenous people was inherited with traditional knowledge on medicinal plants that could inhibit and disintegrate kidney stone formation (Adnan & Othman, 2012; Ong & Norzalina, 1999). Over the years, antiurolithiatic plants had been widely used in the form of decoction, infusion, or juice, to eliminate kidney stones and to prevent their reoccurrence (Ahmed *et al.*, 2018). Previous pre-clinical and clinical studies have established valuable effects of many plants related with kidney functions and stones in experimental animals (Das, Vasudeva & Sharma, 2019).

Basically, the term used in urolithiasis is based on the location of the stone such as nephrolithiasis (renal calculi or kidney stones), ureterolithiasis (ureter calculi

or ureter stone) and cystolithiasis (bladder calculi) (Mikawlawng, Kumar & Vandana, 2014). It is a complex physicochemical process as its formation due to the super saturation of mineral salts in the urinary tract (Balaji, Banji & Banji, 2015). It is estimated to occur in approximately 12% of the global population and its occurrence rate in males is 70-81% and 47-60% in female (Shukla *et al.*, 2017; Soundararajan *et al.*, 2006). It can range from a minor health problem to a life threatening situation since it is a multifactorial disease with high prevalence and reoccurrence rate. There are four categories of urinary stones which are dominantly calcium oxalate (75-90%), followed by struvite (10-15%), uric acid (3-10%) and cysteine stone which only 0.5 to 1% (Aggarwal *et al.*, 2010).

In this study, calcium oxalate (CaOx) stone is highlighted as it is the most common stone and it was prepared experimentally. The properties presence in the extracts of *Anacardium occidentale* (gajus), *Ananus cosmosus* (nanas), *Aquilaria malaccensis* (karas), *Ceiba pentandra* (kekabu), *Centella asiatica* (pegaga), *Euphorbia hirta* (ara tanah), *Ficus carica* (ara), *Melastoma malabathricum* (senduduk), *Piper sarmentosum* (kaduk), and *Tradescantia zebrina* (kura-kura air) were being determined for its antiurolithiatic effects as well as its phytochemical constituents. In evaluation of antiurolithiatic activity, polyherbal drug (cystone) and chemical drug (potassium citrate) were used as a reference standard. The effect of solvent concentration and also extraction time and temperature were evaluated to determine the optimum extraction condition for maximizing antiurolithiatic therapy against CaOx which induced urolithiasis (*in-vitro*) for the selected optimized extracts.

1.2 Problem statement

In recent years, urolithiasis has become a worldwide problem as it is long term ailment that give consequences throughout patient's lifetime (Al-yousofy *et al.*, 2017; Patel *et al.*, 2012). It is generally known as third common affliction of urinary system after urinary tract infection and prostrate disease with estimated occurrence in approximately more than 1/10th of population (Shukla *et al.*, 2017; Bahmani *et al.*, 2016; Hiatt & Friedman, 1982). The prevalence of this ailment has been increasing in the world ranging from 7% to 13% in North America, 5% to 9% in Europe and 1% to 5% in Asia (Liu *et al.*, 2018). In Germany, it is reported that almost 750 000 kidney

stone cases per year with 25% experienced stone reoccurrence (Knoll, 2010; Hesse *et al.*, 2003). This might be influenced by climate, dietary and lifestyle habits (Romero, Akpınar & Assimos, 2010).

Malaysia has shown the same pattern of incidence as it is growing for period of year in 1962 to 1981 per 100 000 populations (Liu *et al.*, 2018; Alatab *et al.*, 2016; Sreenevasan, 1990). Out of more than 4000 people subjected to this disease, the Chinese community lead with of 48%, followed by Malays (37.4%), Indians (13%) and lastly other races with 1.3% (Sreenevasan, 1990). However, another study done in teaching hospital in Kelantan, Malaysia reported that Malay ethnicity dominated with 91.1% (Nouri & Hassali, 2018). There were lack of documentation on this disease in Malaysia and last published was in 1990 (Nouri & Hassali, 2018; Sreenevasan, 1990). In addition, Malaysia has a subtropical climate that could contribute to the formation of kidney stones in humans. Such climate tends to accelerate body dehydration process caused by exposure to hot temperature. This situation causes urine concentration thus leading to stone formation and high frequency of urolithiasis (Hussein *et al.*, 2013).

Nowadays, the management of urolithiasis includes surgical procedures such as extracorporeal shock wave lithotripsy (ESWL), ureteroscopy (URS) and percutaneous nephrolithotomy (PNL). EWSL is the most widely used method as it involves noninvasive procedure that uses sound waves to fragment calculi (Silberstein, Lakin & Parsons, 2008). However, stones that larger than five (5) mm or stones failed to pass through the urinary tract required those interventional procedures (Mikawlawng *et al.*, 2014; Gilhotra, Mohan & Christina, 2013). In addition, this treatment has been proved to have renal side effects as well as quite expensive and high in reoccurrence rate (Tiwari *et al.*, 2012).

Even though there are plentiful of progress in the study of the biological and physical manifestation of urolithiasis, there is truly no satisfactory drug available for the treatment of urolithiasis, particularly for the prevention of reappearance of the stones (Moe, Pearle & Sakhaee, 2011). The pharmaceutical drugs available nowadays are mostly might have adverse effects such as cause nausea, anxiety and also kidney damage that compromise their long-term use (Ankur *et al.*, 2010; Atmani & Khan, 2000). In addition, it was reported that approximately 50% of patients with previous urinary or kidney stones have reappearance within 10 years (Ankur *et al.*, 2010). Therefore, urolithiasis can be considered as a serious disease.

1.3 Research objectives

The main objective of this study is to optimize extraction parameters of selected Malaysian plants on *in-vitro* antiurolithiatic activity. Meanwhile, the specific objectives are as follows:

- i. to assess the best ethnobotanical plant extract that had higher antiurolithiatic activities than standard drugs (positive controls) or no significant different with those drugs ($p>0.05$).
- ii. to determine the optimum extraction parameters for maximum inhibitory and dissolution antiurolithiatic effect on selected plant extracts.
- iii. to evaluate the phytochemical contents and toxicity effect of selected optimized plant extracts.

1.4 Scope of study

In order to achieve the research objective, the scopes of study that has been determined are:

- i. The plant extracts used are *A. occidentale* (leaves), *A. cosmosus* (fruit), *A. malaccensis* (stem), *C. pentandra* (leaves), *C. asiatica* (whole), *E. hirta* (whole), *F. carica* (leaves), *M. malabathricum* (root), *P. sarmentosum* (leaves) and *T. zebrina* (whole).
- ii. The extraction process on preliminary antiurolithiatic assessment extracts was done by decoction method.
- iii. The best plant extracts on screening of antiurolithiatic properties were determined if the plant extracts demonstrate higher or no significant difference with standard drug, ($p>0.05$).
- iv. The antiurolithiatic properties was analyzed by turbidity (nucleation) and titrimetric (calcium oxalate dissolution) assays.
- v. The extraction parameters that were evaluated are solvent concentration, extraction temperature and extraction time.
- vi. The screening of extraction parameters was designated using fractional factorial design (Resolution III, 2^{3-1} design).

- vii. The optimum solvent concentration, extraction temperature and time were determined by using response surface methodology (RSM).
- viii. The phytochemical contents of the optimized extract that were evaluated are phenols, alkaloids, saponins, flavonoids, tannins and terpenoids by qualitative and quantitative methods.
- ix. The toxicity test of the optimized extract was done by using brine shrimp lethality assay.

1.5 Significance of study

The World Health Organization (WHO) reported that around 80% of the population in developing countries still depends on medicinal plants to treat various diseases (Bahmani *et al.*, 2016; Ghatapanadi, Johnson & Rajasab, 2011). Recently, research on beneficial uses of medicinal plants have become mainstream globally including Malaysia. An estimate of 73% of households in Malaysia consume herbal products based on a survey conducted by the Forest Research Institute Malaysia (FRIM) (Ahmad *et al.*, 2015). Malaysia have approximately 15000 species of vascular plants with about 8300 species in Peninsular Malaysia and 12000 species in Sabah and Sarawak (Saw *et al.*, 2010). In addition, around 1300 species out of 14500 flowering plants are recorded to have medicinal use. Same goes to Sabah where about 1200 species out of 7411 species (excluding bryophyta, algae and fungi) found were used statewide for medicinal purposes including kidney stone problems (Kulip *et al.*, 2010; Kulip, 2003).

Besides that, medicinal plants have been used traditionally to treat kidney stone even before the invention of modern treatments (Kumar, Latheef, & Remashree, 2014). There were studies reported that medicinal plants being effective and naturally safe remedies for kidney stone diseases (Alok *et al.*, 2013; Butterweck & Khan, 2009). In the food industry, the formulated herbal supplement drinks from medicinal plants are becoming significant as consumer awareness is increasing in order to develop a healthy lifestyle for managing urolithiasis. Thus, this study will start by screening a few types of Malaysian medicinal plants that have been traditionally reported for its antiurolithiatic potential and determine the optimum conditions for evaluating the antiurolithiatic activities (*in-vitro*).

REFERENCES

- Ab Rahman, S. F. S., Sijam, K., & Omar, D. (2014). Identification and antibacterial activity of phenolic compounds in crude extracts of *Piper sarmentosum* (kadok). *Journal of Pure and Applied Microbiology*, 8, 483–490.
- Abarca-Vargas, R., Peña Malacara, C. F., & Petricevich, V. L. (2016). Characterization of chemical compounds with antioxidant and cytotoxic activities in *bougainvillea x buttiana holttum* and standl, (var. rose) extracts. *Antioxidants*, 5(45), 1–11.
- Abascal, K., Ganora, L., & Yarnell, E. (2005). The effect of freeze-drying and its implications for botanical medicine: A review. *Phytotherapy Research*, 19(September 2004), 655–660.
- Abdullah, N., & Chin, N. L. (2010). Simplex-centroid mixture formulation for optimised composting of kitchen waste. *Bioresource Technology*, 101(21), 8205–8210.
- Abouelela, M. E., Orabi, M. A. A., Abdelhamid, R. A., Abdelkader, M. S. A., & Darwish, F. M. M. (2018). Chemical and cytotoxic investigation of non-polar extract from *Ceiba pentandra* (L.) Gaertn.: A study supported by computer based screening. *Journal of Applied Pharmaceutical Science*, 8(7), 57–64.
- Abu-reidah, I. M., Ali-shtayeh, M. S., Jamous, R. M., Arráez-román, D., & Segura-carretero, A. (2015). HPLC–DAD–ESI-MS/MS screening of bioactive components from *Rhus coriaria* L. (Sumac) fruits. *Food Chemistry*, 166, 179–191.
- Abu Bakar, F. I., Abu Bakar, M. F., Abdullah, N., Endrini, S., & Fatmawati, S. (2020). Optimization of extraction conditions of phytochemical compounds and anti-gout activity of *Euphorbia hirta* L. (ara tanah) using response surface methodology and liquid chromatography-mass spectrometry (LC-MS) analysis. *Evidence-Based Complementary and Alternative Medicine*, 2020, 1–13.
- Abu Bakar, M. F., Mohamed, M., Rahmat, A., & Fry, J. (2009). Phytochemicals and



antioxidant activity of different parts of bambangan (*Mangifera pajang*) and tarap (*Artocarpus odoratissimus*). *Food Chemistry*, 113, 479–483.

Adnan, N., & Othman, N. (2012). The relationship between plants and the Malay culture. *Procedia - Social and Behavioral Sciences*, 42(2012), 231–241.

Agarwal, K., & Varma, R. (2015). In-vitro calcium oxalate crystallization inhibition by *Achyranthes aspera* L. and *Bryophyllum pinnatum* Lam. *British Journal of Pharmaceutical Research*, 5(2), 146–152.

Aggarwal, A., Tandon, S., Singla, S. K., & Tandon, C. (2010). Diminution of oxalate induced renal tubular epithelial cell injury and inhibition of calcium oxalate crystallization in vitro by aqueous extract of *Tribulus terrestris*. *International Brazilian Journal of Urology*, 36(4), 480–488.

Aggarwal, K. P., Narula, S., Kakkar, M., & Tandon, C. (2013). Nephrolithiasis: Molecular mechanism of renal stone formation and the critical role played by modulators. *BioMed Research International*, 2013, 1–21.

Ahmad, Farizah, Zaidi, M. A. S., Sulaiman, N., & Majid, F. A. A. (2015). Issues and challenges in the development of the herbal industry in Malaysia. *Preceding PERKEM*, 10(September), 227–238.

Ahmad, Fasihuddin, & Ismail, G. (2003). Medicinal plants used by Kadazandusun communities around Crocker Range. *ASEAN Review of Biodiversity and Environmental Conversation (ARBEC)*, 1, 1–10. Retrieved from

Ahmaed, D. T., Mohammed, M., Masaad, A. M., & Tajuddin, S. N. (2017). Investigation of agarwood compounds in *Aquilaria malaccensis* & *Aquilaria rostrata* chipwood by using solid phase microextraction. *Biomedical Journal of Scientific & Technical Research*, 1(6), 1609–1616.

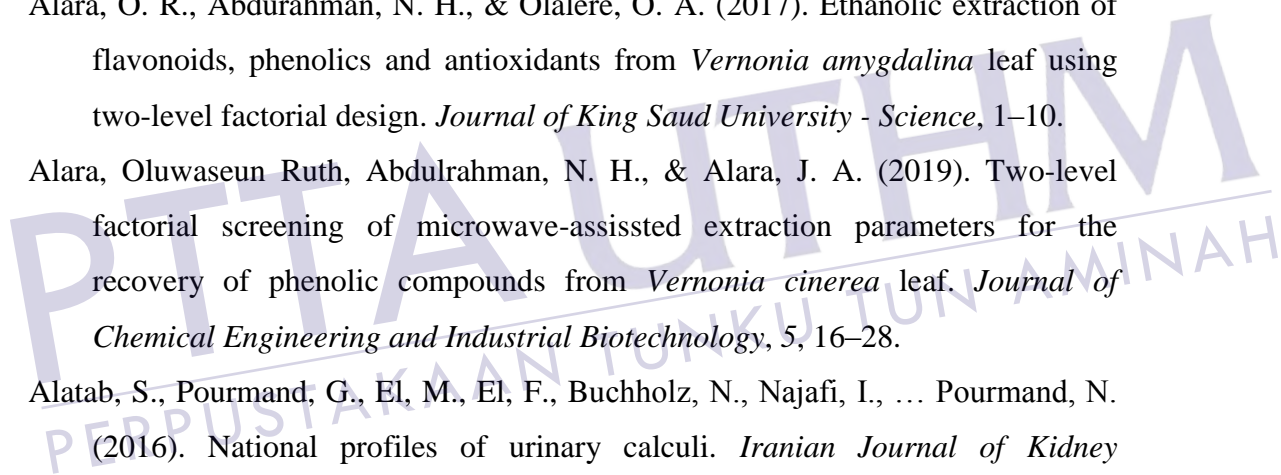
Ahmed, M., Ji, M., Qin, P., Gu, Z., Liu, Y., Sikandar, A., ... Javeed, A. (2019). Phytochemical screening, total phenolic and flavonoids contents and antioxidant activities of *Citrullus colocynthis* L. and *Cannabis sativa* L. *Applied Ecology and Environmental Research*, 17(3), 6961–6979.

Ahmed, S., Hasan, M. M., & Alam, Z. (2016). In-vitro urolithiasis models: An evaluation of prophylactic management against kidney stones. *Journal of Pharmacognosy and Phytochemistry*, 5(3), 28–35.

Ahmed, S., Hasan, M. M., & Mahmood, Z. A. (2016). Antiurolithiatic plants: Multidimensional pharmacology. *Journal of Pharmacognosy and Phytochemistry*, 5(2), 04–24.



- Ahmed, S., Mohtasheemul, M., Khan, H., & Alam, Z. (2018). The mechanistic insight of polyphenols in calcium oxalate urolithiasis mitigation. *Biomedicine & Pharmacotherapy*, 106(June), 1292–1299.
- Akhtar, S. S., Mular, S. M., Khan, N. D., Khan, Z. H., & Sohail, S. (2017). In-vitro study of aqueous leaf extract of *Raphanus sativus* var. for inhibition of calcium oxalate crystallization. *Bioscience Discovery*, 8(2), 153–157.
- Al-Snafi, A. E. (2017). Pharmacology and therapeutic potential of *Euphorbia hirta* (Syn: *Euphorbia pilulifera*)- A review. *IOSR Journal of Pharmacy (IOSRPHR)*, 07(03), 07–20.
- Al-yousofy, F., Gumaih, H., Ibrahim, H., & Alasbahy, A. (2017). Parsley! Mechanism as antiurolithiasis remedy. *International Journal of Medical and Health Research*, 3(7), 35–40.
- Alara, O. R., Abdurahman, N. H., & Olalere, O. A. (2017). Ethanolic extraction of flavonoids, phenolics and antioxidants from *Vernonia amygdalina* leaf using two-level factorial design. *Journal of King Saud University - Science*, 1–10.
- Alara, Oluwaseun Ruth, Abdurahman, N. H., & Alara, J. A. (2019). Two-level factorial screening of microwave-assisted extraction parameters for the recovery of phenolic compounds from *Vernonia cinerea* leaf. *Journal of Chemical Engineering and Industrial Biotechnology*, 5, 16–28.
- Alatab, S., Pourmand, G., El, M., El, F., Buchholz, N., Najafi, I., ... Pourmand, N. (2016). National profiles of urinary calculi. *Iranian Journal of Kidney Diseases*, 10(2), 51–61.
- Alberti, A., Zielinski, A. A. F., Zardo, D. M., Demiate, I. M., Nogueira, A., & Mafra, L. I. (2014). Optimisation of the extraction of phenolic compounds from apples using response surface methodology. *Food Chemistry*, 149, 151–158.
- Alok, S., Jain, S. K., Verma, A., Kumar, M., & Sabharwal, M. (2013). Pathophysiology of kidney, gallbladder and urinary stones treatment with herbal and allopathic medicine: A review. *Asian Pacific Journal of Tropical Disease*, 3(6), 496–504.
- Altemimi, A., Lakhssassi, N., Baharlouei, A., Watson, D. G., & Lightfoot, D. A. (2017). Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant extracts. *Plants*, 6(42), 1–23.
- Altıok, E., Bayçin, D., Bayraktar, O., & Ülkü, S. (2008). Isolation of polyphenols from the extracts of olive leaves (*Olea europaea* L.) by adsorption on silk



- fibroin. *Separation and Purification Technology*, 62(2), 342–348.
- Ankur, C., Amarchand, P., Aadarsh, C., Deepa, I., R.S, P., & U.K, P. (2010). Potential of medicinal plants in kidney, gall and urinary stones. *International Journal of Drug Development & Research*, 2(2), 431–447.
- Anwar, F., Kalsoom, U., Sultana, B., Mushtaq, M., Mehmood, T., & Arshad, H. A. (2013). Effect of drying method and extraction solvent on the total phenolics and antioxidant activity of cauliflower (*Brassica oleracea* L.) extracts. *International Food Research Journal*, 20(2), 653–659.
- Arya, P., Pandey, S., & Verma, V. (2017). Kidney stones formation and use of medicinal plants as antiurolithiatic agents. *Universal Journal of Pharmaceutical Research*, 2(4), 42–48.
- Asaduzzaman, M., Rana, S., Hasan, S. M. R., Hossain, M., & Das, N. (2015). Cytotoxic (brine shrimp lethality bioassay) and antioxidant investigation of *Barringtonia acutangula* (L.). *International Journal of Pharma Sciences and Research*, 6(8), 1179–1185.
- Asha, S., Deevika, B., & Sadiq, A. M. (2014). *Euphorbia hirta* Linn-a review on traditional uses, phytochemistry and pharmacology. *World Journal of Pharmaceutical Research*, 3(4), 180–205.
- Aslam, M. S., Ahmad, M. S., Ahmad, M. A., & Akhlaq, M. (2017). An updated review on phytochemical and pharmacological properties of *Piper sarmentosum*. *Current Trends in Biotechnology and Pharmacy*, 11(4), 345–356.
- Atmani, F., & Khan, S. R. (2000). Effects of an extract from *Herniaria hirsuta* on calcium oxalate crystallization in vitro. *BJU International*, 85, 621–625.
- Atodariya, U., Barad, R., Upadhyay, S., & Upadhyay, U. (2013). Anti-urolithiatic activity of *Dolichos biflorus* seeds. *Journal of Pharmacognosy and Phytochemistry*, 2(2), 209–213.
- Aybaster, Ö., Is, E., & Saliha, S. (2013). Optimization of ultrasonic-assisted extraction of antioxidant compounds from blackberry leaves using response surface methodology. *Industrial Crops and Products*, 44, 558–565.
- Azahar, N. F., Gani, S. S. A., & Mohd Mokhtar, N. F. (2017). Optimization of phenolics and flavonoids extraction conditions of *Curcuma Zedoaria* leaves using response surface methodology. *Chemistry Central Journal*, 11(1), 1–10.
- Azwanida, N. N. (2015). A review on the extraction methods use in medicinal plants, principle, strength and limitation. *Medicinal & Aromatic Plants*, 4(3),



PTTA UTHM
PUSAT PENYELIDIKAN TUMBUHAN
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3–8.

- Badgujar, S. B., Patel, V. V., Bandivdekar, A. H., & Mahajan, R. T. (2014). Traditional uses, phytochemistry and pharmacology of *Ficus carica*: A review. *Pharmaceutical Biology*, 52(11), 1487–1503.
- Bahmani, M., Baharvand-Ahmadi, B., Tajeddini, P., Rafieian-Kopaei, M., & Naghdi, N. (2016). Identification of medicinal plants for the treatment of kidney and urinary stones. *Journal of Renal Injury Prevention*, 5(3), 129–133.
- Balaji, L. G., Banji, D., & Banji, O. J. F. (2015). Evaluation of antiurolithiatic activity of the aqueous and alcoholic extracts of roots of *Boerhaavia diffusa*. *Indo American Journal of Pharmaceutical Research*, 5(01), 525–530.
- Bandar, H., Hijazi, A., Rammal, H., Hachem, A., Saad, Z., & Badran, B. (2013). Techniques for the extraction of bioactive compounds from Lebanese *Urtica dioica*. *American Journal of Phytomedicine and Clinical Therapeutics*, 1(6), 3158–3168.
- Banerjee, S., & Chatterjee, J. (2015). Efficient extraction strategies of tea (*Camellia sinensis*) biomolecules. *Journal of Food Science and Technology*, 52(6), 3158–3168.
- Barros, M. E., Lima, R., Mercuri, L. P., Matos, J. R., Schor, N., & Boim, M. A. (2006). Effect of extract of *Phyllanthus niruri* on crystal deposition in experimental urolithiasis. *Urological Research*, 34(6), 351–357.
- Basavaraj, D. R., Biyani, C. S., Browning, A. J., & Cartledge, J. J. (2007). The role of urinary kidney stone inhibitors and promoters in the pathogenesis of calcium containing renal stones. *EAU-EBU Update Series*, 5(3), 126–136.
- Batubara, R., Hanum, T. I., & Surjanto. (2018). Phytochemical and tannin content in two species of agarwood leaves from Mandailing Natal Regency North Sumatera Province. *AIP Conference Proceedings*, 2049(December), 1–4.
- Bawari, S., Negi Sah, A., & Tewari, D. (2018). Antiurolithiatic activity of *Daucus carota*: an in-vitro study. *Pharmacognosy Journal*, 10(5), 880–884.
- Behera, S. K., Meena, H., Chakraborty, S., & Meikap, B. C. (2018). Application of response surface methodology (RSM) for optimization of leaching parameters for ash reduction from low-grade coal. *International Journal of Mining Science and Technology*, 28(4), 621–629.
- Behravan, E., Heidari, M. R., Heidari, M., Fatemi, G., Etemad, L., Taghipour, G., & Abbasifard, M. (2012). Comparison of gastric ulcerogenicity of percolated extract of *Anacardium occidentale* (cashew nut) with indomethacin in rats.



Pakistan Journal of Pharmaceutical Science, 25(1), 111–115.

- Belwal, T., Dhyani, P., Bhatt, I. D., Rawal, R. S., & Pande, V. (2016). Optimization extraction conditions for improving phenolic content and antioxidant activity in *Berberis asiatica* fruits using response surface methodology (RSM). *Food Chemistry*, 207(June), 115–124.
- Benalia, H., Djeridane, A., Bensafieddine, F., & Yousfi, M. (2016). High in vitro antiurolithiatic effect of *Pituranthos scoparius* roots extracts. *Pharmacologyonline*, 1, 31–43.
- Bergman, M. E., Davis, B., & Phillips, M. A. (2019). Medically useful plant terpenoids: biosynthesis, occurrence, and mechanism of action. *Molecules*, 24, 3961.
- Betanabhatla, K. S., Christina, A. J. M., Sundar, B. S., Selvakumar, S., & Saravanan, K. S. (2009). Antilithiatic activity of *Hibiscus sabdariffa* Linn. on ethylene glycol-induced lithiasis in rats. *Natural Product Radiance*, 8(1), 43–47.
- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., & Escalera, L. A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76(5), 965–977.
- Biglarkhani, M., Zargar, M. A. A., Hashem-Dabaghian, F., Behbahani, F. A., Meyari, A., & Sadeghpour, O. (2017). Common antiurolithiatic medicinal plants in Persian medicine: A review. *Indo American Journal of Pharmaceutical Sciences*, 4(12), 4838–4846.
- Bijarnia, R. K., Kaur, T., Singla, S. K., & Tandon, C. (2010). Non-surgical management therapies for kidney stones. *Journal of Pharmaceutical Education and Research*, 1(1), 21–25.
- Bucić-Kojić, A., Planinić, M., Tomas, S., Jakobek, L., & Šeruga, M. (2009). Influence of solvent and temperature on extraction of phenolic compounds from grape seed, antioxidant activity and colour of extract. *International Journal of Food Science and Technology*, 44(12), 2394–2401.
- Burda, S., Ozleszek, W., & Lee, C. Y. (1990). Phenolic compounds and their changes in apples during maturation and cold storage. *Journal of Agricultural and Food Chemistry*, 30, 945–948.
- Butterweck, V., & Khan, S. R. (2009). Herbal medicines in the management of urolithiasis: Alternative or complementary? *Planta Medica*, 75(10), 1095–1103.



PTTA (Pusat Teknologi dan Teknologi Akademik)
PEPPUSIAKAN PUSAT KUTIPAMINAH

- Byahatti, V. V, Pai, K. V., & Souza, M. G. D. (2010). Effect of phenolic compounds from *Bergenia ciliata* (Haw.) Sternb. leaves on experimental kidney stones. *Ancient Science of Life*, 30(1), 14–17.
- Calixto, J. B., Yunes, R. A., Neto, A. S., Valle, R. M., & Rae, G. A. (1984). Antispasmodic effects of an alkaloid extracted from *Phyllanthus sellowianus*: a comparative study with papaverine. *Brazilian Journal of Medical and Biological Research*, 17(3–4), 313–321.
- Carvalho, M. (2018). Urinary pH in calcium oxalate stone formers: does it matter? *Brazilian Journal of Nephrology*, 40(1), 6–7.
- Chai, T.-T., Wong, F.-C., Manan, F. A., Ooh, K.-F., & Ismail, N. I. M. (2014). *Orthosiphon aristatus*: A review of traditional uses, phytochemical profile, and pharmacological properties. *Traditional and Folk Herbal Medicine: Recent Researches*, 2, 153–187.
- Chan, E. W. C., Lim, Y. Y., Wong, S. K., Lim, K. K., Tan, S. P., Lianto, F. S., & Yong, M. Y. (2009). Effects of different drying methods on the antioxidant properties of leaves and tea of ginger species. *Food Chemistry*, 113(1), 166–172.
- Chan, Eric Wei Chiang, Baba, S., Chan, H. T., Kainuma, M., Inoue, T., & Wong, S. K. (2017). Ulam herbs: A review on the medicinal properties of *Anacardium occidentale* and *Barringtonia racemosa*. *Journal of Applied Pharmaceutical Science*, 7(2), 241–247.
- Chaudhary, A., Singla, S. K., & Tandon, C. (2010). In-vitro evaluation of *Terminalia arjuna* on calcium phosphate and calcium oxalate crystallization. *Indian Journal of Pharmaceutical Sciences*, 72(3), 340–345.
- Chaudhary, V., Kumar, V., Sunil, Vaishali, Singh, K., Kumar, R., & Kumar, V. (2019). Pineapple (*Ananas cosmosus*) product processing: A review. *Journal of Pharmacognosy and Phytochemistry*, 8(3), 4642–4652.
- Che Sulaiman, I. S., Basri, M., Fard Masoumi, H. R., Chee, W. J., Ashari, S. E., & Ismail, M. (2017). Effects of temperature, time, and solvent ratio on the extraction of phenolic compounds and the anti-radical activity of *Clinacanthus nutans* Lindau leaves by response surface methodology. *Chemistry Central Journal*, 11(1), 1–11.
- Cheah, S. Y., Magdalene, C. Y., Eldwin Lim, C. Z., Wong, M. H., Amir, S., Daniel, S., ... Chin, J. H. (2017). In-vitro antioxidant and acetylcholinesterase



- inhibitory activities of *Tradescantia zebrina*. *Research Journal of Pharmaceutical , Biological and Chemical Sciences*, 8(1), 82–87.
- Chen, M., Weng, S., Hsu, C., Lin, H., Su, S., & Wang, J. (2016). Urolithiasis risk: a comparison between healthcare providers and the general population. *BMC Health Services Research*, 16(1), 273–279.
- Chieng, T., Assim, Z., & Fasihuddin, B. (2008). Toxicity and antitermite activities of the essential oils from *Piper sarmentosum*. *The Malaysian Journal of Analytical Sciences*, 12(1), 234–239.
- Chinnala, K. M., Shanigarm, S., & Elsani, M. M. (2013). Antiuro lithiatic activity of the plant extracts of *Solanum virginianum* on ethylene glycol induced urolithiasis in rats. *International Journal of Pharmacy and Biological Science*, 3(4), 328–334.
- Chisom, I. F., Chukwu, O. N., & Okeke, C. U. (2014). Comparative phytochemical and proximate analyses on *Ceiba pentandra* (L) Gaertn. and *Bombax buonopozense* (P) Beauv. *International Journal of Herbal Medicine*, 2(2), 162–167.
- Choubey, A., Choubey, A., Jain, P., Iyer, D., & Patil, U. K. (2010). Assessment of *Ceiba pentandra* on calcium oxalate urolithiasis in rats. *Der Pharma Chemica*, 2(6), 144–156.
- Coe, F. G., Parikh, D. M., & Johnson, C. A. (2010). Alkaloid presence and brine shrimp (*Artemia salina*) bioassay of medicinal species of eastern Nicaragua. *Pharmaceutical Biology*, 48(4), 439–445.
- Corradini, E., Fogliaa, P., Giansantia, P., Gubbiottia, R., Samperi, R., & Aldo, L. (2011). Flavonoids: Chemical properties and analytical methodologies of identification and quantitation in foods and plants. *Natural Product Research*, 25(5), 469–495.
- Czitrom, V. (1999). One-factor-at-a-time versus designed experiments. *American Statistician*, 53(2), 126–131.
- Dai, J., & Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15, 7313–7352.
- Danladi, S., Wan-Azemin, A., Sani, Y. N., Mohd, K. S., Us, M. R., Mansor, S. M., & Dharmaraj, S. (2015). Phytochemical screening, antioxidant potential and cytotoxic activity of *Melastoma malabathricum* Linn. from different locations. *International Journal of Pharmacy and Pharmaceutical Sciences*, 7(7), 408–



413.

- Das, S., Vasudeva, N., & Sharma, S. (2019). Kidney disorders and management through herbs: A Review. *The Journal of Phytopharmacology*, 8(1), 21–27.
- Dash, G. K., Swe, M., & Mathews, A. (2017). *Tradescantia zebrina*: A promising medicinal plant. *Indo American Journal of Pharmaceutical Sciences*, 4(10), 3498–3502.
- Dash, M., Patra, J. K., & Panda, P. P. (2008). Phytochemical and antimicrobial screening of extracts of *Aquilaria agallocha* Roxb. *African Journal of Biotechnology*, 7(20), 3531–3534.
- Defilipps, R. A., & Krupnick, G. A. (2018). The medicinal plants of Myanmar. *PhotoKeys*, 341, 1–341.
- Dent, M., Dragović-Uzelac, V., Penić, M., Brčić, M., Bosiljkov, T., & Levaj, B. (2013). The effect of extraction solvents, temperature and time on the composition and mass fraction of polyphenols in dalmatian wild sage (*Salvia officinalis* L.) extracts. *Food Technology and Biotechnology*, 51(1), 84–91.
- Desai, D., Raorane, C., Patil, S., Gadgil, R., & Patkar, D. (2017). *Anacardium occidentale*: Fountain of phytochemicals; the qualitative profiling. *World Journal of Pharmaceutical Research*, 6(5), 585–592.
- Desai, S. D., Desai, D. G., & Kaur, H. (2009). Saponins and their biological activities. *Pharma Times*, 41(3), 13–16.
- Devi, S., Jahangir, R., & Kumar, M. (2017). Phytochemical screening of whole plant extract of *Euphorbia hirta* L. *International Journal of Biology Reserch*, 2(4), 91–93.
- Devkar, R. A., Chaudhary, S., Adepu, S., Xavier, S. K., Chandrashekar, K. S., & Setty, M. M. (2016). Evaluation of antiurolithiatic and antioxidant potential of *Lepidagathis prostrata*: A Pashanbhed plant. *Pharmaceutical Biology*, 54(7), 1237–1245.
- Dewanto, V., Wu, X., Adom, K. K., & Liu, R. H. (2002). Thermal processing enhances the nutritional value of tomatoes by increasing total antioxidant activity. *Journal of Agricultural and Food Chemistry*, 50, 3010–3014.
- Dibazar, R. (2014). *Optimization of ultrasound-assisted extraction of anthocyanins from Nova Scotian lowbush blueberries (Vaccinium angustifolium Aiton L.)*. Dalhousie University, Halifax, Nova Scotia.
- Dibazar, R., Bonat Celli, G., Brooks, M. S. L., & Ghanem, A. (2015). Optimization



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PERPUSTAKAAN TUN AMINAH

- of ultrasound-assisted extraction of anthocyanins from lowbush blueberries (*Vaccinium angustifolium* Aiton). *Journal of Berry Research*, 5(3), 173–181.
- Dinnimath, B. M., Jalalpure, S. S., & Patil, U. K. (2017). Antiurolithiatic activity of natural constituents isolated from *Aerva lanata*. *Journal of Ayurveda and Integrative Medicine*, 8(4), 226–232.
- Doss, V. A., & Thangavel, K. P. (2011). Antioxidant and antimicrobial activity using different extracts of *Anacardium occidentale* L. *International Journal of Applied Biology and Pharmaceutical Technology*, 2(3), 436–443.
- El Aziz, M. M. A., Ashour, A. S., & Gomha Melad, A. S. (2019). A review on saponins from medicinal plants: chemistry, isolation, and determination. *Journal of Nanomedicine Research*, 7(4), 282–288.
- Elboughdiri, N. (2018). Effect of time, solvent-solid ratio, ethanol concentration and temperature on extraction yield of phenolic compounds from olive leaves. *Engineering, Technology and Applied Science Research*, 8(2), 2805–2808.
- Elias, M. F., Ibrahim, H., & Mahamod, W. R. W. (2017). A review on the Malaysian *Aquilaria* species in karas plantation and agarwood production. *International Journal of Academic Research in Business and Social Sciences*, 7(4), 1021–1029.
- Erickson, S. B., Vrtiska, T. J., Canzanello, V. J., & Lieske, J. C. (2010). Cystone® for 1 year did not change urine chemistry or decrease stone burden in cystine stone formers. *Urological Research*, 39(3), 197–203.
- Erickson, S. B., Vrtiska, T. J., & Lieske, J. C. (2011). Effect of Cystone® on urinary composition and stone formation over a one year period. *Phytomedicine*, 18(10), 863–867.
- Eweka, A. O., & Enogieru, A. (2011). Effects of oral administration of *Phyllanthus amarus* leaf extract on the kidneys of adult wistar rats-a histological study. *African Journal of Traditional, Complementary and Alternative Medicines*, 8(3), 307–311.
- Ezeonu, C. S., & Ejikeme, C. M. (2016). Qualitative and quantitative determination of phytochemical contents of indigenous Nigerian softwoods. *New Journal of Science*, 2016, 1–9.
- Farook, N. A. M., Rajesh, S., & Jamuna, M. (2009). Inhibition of mineralization of urinary stone forming minerals by medicinal-plants. *Journal of Chemistry*, 6(3), 938–942.



- Fitzhugh, D. J., Shan, S., Dewhirst, M. W., & Hale, L. P. (2008). Bromelain treatment decreases neutrophil migration to sites of inflammation. *Clinical Immunology*, 128(1), 66–74.
- Frassetto, L., & Kohlstadt, I. (2011). Treatment and prevention of kidney stones: An update. *American Family Physician*, 84(11), 1234–1242.
- Gadir, S. A. (2012). Assessment of bioactivity of some Sudanese medicinal plants using brine shrimp (*Artemia salina*) lethality assay. *Journal of Chemical and Pharmaceutical Research*, 4(12), 5145–5148.
- George, M., Joseph, L., & Ramaswamy. (2009). Anti-allergic, anti-pruritic, and anti-inflammatory activities of *Centella asiatica* extracts. *African Journal of Traditional, Complementary and Alternative Medicines*, 6(4), 554–559.
- Ghatapanadi, S. R., Johnson, N., & Rajasab, A. H. (2011). Documentation of folk knowledge on medicinal plants of Gulbarga district, Karnataka. *Indian Journal of Traditional Knowledge*, 10(2), 349–353.
- Ghazali, N. F., Mohd, M. A., Ibrahim, M. A., & Muhammad, T. S. T. (2018). Phytochemical and pharmacological profile of Kaduk (*Piper sarmentosum* Roxb.). *Malayan Nature Journal*, 70(2), 195–202.
- Ghorai, N., Chakraborty, S., Guchait, S., Saha, S. K., & Biswas, S. (2012). Estimation of total terpenoids concentration in plant tissues using a monoterpene, Linalool as standard reagent. *Protocol Exchange*, 5(10), 1–6.
- Gilhotra, U. K., Mohan, G., & Christina, A. J. M. (2013). Antilithiatic activity of poly-herbal formulation tablets by in-vitro method. *Journal of Applied Pharmaceutical Science*, 3(5), 43–48.
- Gopalakrishnan, K., & Udayakumar, R. (2017). Phytochemical content of leaf and stem of *Marsilea quadrifolia* (L.). *Journal of Plant Science and Phytopathology*, 1, 26–37.
- Goswami, P. K., Srivastava, R. S., Samant, M., & Khale, A. (2013). Urolithiasis : an overview. *International Journal of Pharmaceutical & Biological Archive*, 4(6), 1119–1122.
- Hamid, A. A., Shah, Z. M., Muse, R., & Mohamed, S. (2002). Characterisation of antioxidative activities of various extracts of *Centella asiatica* (L.) Urban. *Food Chemistry*, 77(4), 465–469.
- Hamidi, M. R., Jovanova, B., & Panovska, T. K. (2014). Toxicological evaluation of the plant products using Brine shrimp (*Artemia salina* L.) model. *Macedonian*



Pharmaceutical Bulletin, 60(1), 9–18.

- Han, H., Segal, A. M., Seifter, J. L., & Dwyer, J. T. (2015). Nutritional management of kidney stones (nephrolithiasis). *Clinical Nutrition Research*, 4(3), 137–152.
- Hashim, Y. Z. H. Y., Kerr, P. G., Abbas, P., & Mohd Salleh, H. (2016). *Aquilaria* spp. (agarwood) as source of health beneficial compounds: A review of traditional use, phytochemistry and pharmacology. *Journal of Ethnopharmacology*, 189, 331–360.
- He, Q., Du, B., & Xu, B. (2018). Extraction optimization of phenolics and antioxidants from black goji berry by accelerated solvent extractor using response surface methodology. *Applied Sciences (Switzerland)*, 8(10), 1905.
- Hendra, H., Moeljopawiro, S., & Nuringtyas, T. R. (2016). Antioxidant and antibacterial activities of agarwood (*Aquilaria malaccensis* Lamk.) leaves. *AIP Conference Proceedings*, 1755(July 2016), 1–9.
- Heras, B. De, Rodríguez, B., Boscá, L., & Villar, A. M. (2003). Terpenoids: sources, structure elucidation and therapeutic potential in inflammation. *Current Topics in Medicinal Chemistry*, 3, 53–67.
- Hesse, A., Brändle, E., Wilbert, D., Köhrmann, K. U., & Alken, P. (2003). Study on the prevalence and incidence of urolithiasis in Germany comparing the years 1979 vs. 2000. *European Urology*, 44(6), 709–713.
- Hiatt, R. A., & Friedman, G. D. (1982). The frequency of kidney and urinary tract diseases in a defined population. *Kidney International*, 22(1982), 63–68.
- Hossain, M. A., Al-Mijizy, Z. H., Al-Rashidi, K. K., Weli, A. M., & Al-Riyami, Q. (2013). Effect of temperature and extraction process on antioxidant activity of various leaves crude extracts of *Thymus vulgaris*. *Journal of Coastal Life Medicine*, 1(2), 130–134.
- Huang, L., Chen, S., & Yang, M. (2012). *Euphorbia hirta* (Feiyangcao): A review on its ethnopharmacology, phytochemistry and pharmacology. *Journal of Medicinal Plants Research*, 6(39), 5176–5185.
- Hussain, K., Hashmi, F. K., Latif, A., Ismail, Z., & Sadikun, A. (2012). A review of the literature and latest advances in research of *Piper sarmentosum*. *Pharmaceutical Biology*, 50(8), 1045–1052.
- Hussein, N., Sadiq, S., Kamaliah, M., NorAkmal, A., & Gohar, M. (2013). Twenty-four-hour urine constituents in stone formers: A study from the northeast part of Peninsular Malaysia. *Saudi Journal of Kidney Diseases and Transplantation*,



24(3), 630–637.

- Ibrahim, R., Abubakar, E. M., Misau, M. S., & Lamarin, B. G. (2017). Percentage yield and acute toxicity of the plant extracts of *Ceiba pentandra* grown in Bauchi State, North Eastern Nigeria. *Journal of Pharmacognosy and Phytochemistry*, 6(5), 1777–1779.
- Ilaiyaraja, N., Likhith, K. R., Sharath Babu, G. R., & Khanum, F. (2015). Optimisation of extraction of bioactive compounds from *Feronia limonia* (wood apple) fruit using response surface methodology (RSM). *Food Chemistry*, 173, 348–354.
- Itam, A., Ismail, Z., & Majid, A. M. S. A. (2009). In vitro studies on calcium oxalate crystal growth inhibition of *Strobilanthes crispus* extracts. *Jurnal Riset Kimia*, 3(1), 8.
- Iyare, G. I., Omorodion, N. T., Erameh, T. O., Ackhukwu, P. U., & Ogochukwu, A. G. (2017). The effects of *Anacardium occidentale* leave extract on histology of selected organs of Wistar rats. *MOJ Biology and Medicine*, 2(2), 216–221.
- Jain, A., Roshnibala, S., Kanjilal, P. B., Singh, R. S., & Singh, H. B. (2007). Aquatic/semi-aquatic plants used in herbal remedies in the wetlands of Manipur, Northeastern India. *Indian Journal of Traditional Knowledge*, 6(2), 346–351.
- Jain, S. K. (2003). Notable foreign medicinal uses for some plants of Indian tradition. *Indian Journal of Traditional Knowledge*, 2(4), 321–332.
- Jaiswal, H., Singh, O. J., Chauhan, A., Sahu, M. K., & Dv, S. P. (2018). A review on tannins. *European Journal of Biotechnology and Bioscience*, 6(3), 16–17.
- Jamal, J. A., Ghafar, Z. A., & Husain, K. (2011). Medicinal plants used for postnatal care in Malay traditional medicine in the Peninsular Malaysia. *Pharmacognosy Journal*, 3(24), 15–24.
- James, J. T., & Dubery, I. A. (2009). Pentacyclic triterpenoids from the medicinal herb, *Centella asiatica* (L.) Urban. *Molecules*, 14, 3922–3941.
- Jamil, S. S., Nizami, Q., & Salam, M. (2007). *Centella asiatica* (Linn.) Urban: a review. *Natural Product Radiance*, 6(2), 158–170.
- Jaradat, N. A. (2005). Medical plants utilized in Palestinian folk medicine for treatment of diabetes mellitus and cardiac diseases. *Journal of Al-Aqsa University*, 9, 1–28.
- Jayaraman, U. C., & Gurusamy, A. (2018). Review on uro-lithiasis pathophysiology



- and Aesculapian discussion. *IOSR Journal of Pharmacy*, 8(2), 30–42.
- Jenitha, X. A., & Anusuya, A. (2016). Phytochemical screening and in vitro antioxidant activity of *Ananas comosus*. *International Journal of Research in Pharmacology and Pharmacotherapeutics*, 5(2), 162–169.
- Jha, R., Tahil Ramani, P., Patel, D., Desai, S., Meshram, D., & Rahul Jha, C. (2016). Phytochemical analysis and in-vitro urolithiatic activity of *Peltophorum pterocarpum* leaves (DC) Baker. *Journal of Medicinal Plants Studies*, 18(43), 18–22.
- Joffry, S. M., Yob, N. J., Rofiee, M. S., Affandi, M. M. R. M. M., Suhaili, Z., Othman, F., ... Zakaria, Z. A. (2012). *Melastoma malabathricum* (L.) Smith ethnomedicinal uses, chemical constituents, and pharmacological properties: a review. *Evidence-Based Complementary and Alternative Medicine*, 2012, 1–48.
- Jordan, M. G., Disney, B., Armstrong, E. F., & Luther, L. J. (2016). Dissolution of canine uroliths by Moringha root extracts using various aqueous and organic solvents. *International Journal of Veterinary & Wildlife Sciences*, 1(1), 1–5.
- Jourshabani, M., Badiei, A., Lashgari, N., & Mohammadi Ziarani, G. (2016). Application of response surface methodology as an efficient approach for optimization of operational variables in benzene hydroxylation to phenol by V/SBA-16 nanoporous catalyst. *Journal of Nanostructures*, 6(2), 105–113.
- Joy, J., & Jayachandran, T. P. (2018). Phytochemical and in vitro antiurolithiatic activity of Thlaspi Bursa Pastoris Mother Tincture. *Scholars Academic Journal of Pharmacy*, 7(7), 348–353.
- Kamaludin, N. H. I., & Jaafar, N. S. A. (2017). Evaluation of antioxidant activity from different plant parts of senduduk herb: extraction conditions optimization of selected plant part. *MATEC Web of Conferences*, 97, 1–10.
- Kamanja, I. T., Mbaria, J. M., Gathumbi, P. K., Mathiu, M., & Kiama, S. (2018). Cytotoxicity of selected medicinal plants extracts using the brine shrimp lethality assay from Samburu county, Kenya. *The Journal of Medical Research*, 4(5), 249–255.
- Kamarudin, N. A., Markom, M., & Latip, J. (2016). Effects of solvents and extraction methods on herbal plants *Phyllanthus niruri*, *Orthosiphon stamineus* and *Labisia pumila*. *Indian Journal of Science and Technology*, 9(21), 1–5.
- Kausar, J., Muthumani, D., Hedina, A., Sivasamy, & Anand, V. (2016). Review of the phytochemical and pharmacological activities of *Euphorbia hirta* Linn.



Pharmacognosy Journal, 8(4), 310–313.

- Kaushik, J., & Kundu, N. (2018). Phytochemical screening, anti-oxidant and anti-microbial activity of polyphenolic flavonoids isolated from fruit of *Ananas comosus* in various solvents. *International Journal of Scientific and Research Publications*, 8(2), 31–55.
- Khan, R. A., Khan, M. R., & Sahreen, S. (2010). Evaluation of *Launaea procumbens* use in renal disorders: A rat model. *Journal of Ethnopharmacology*, 128(2), 452–461.
- Khanbabaee, K., & Ree, T. Van. (2001). Tannins: classification and definition. *Natural Product Reports*, 18, 641–649.
- Khare, P., Mishra, V. K., K., A., Bais, N., & Singh, R. (2014). Study on in-vitro antilithiatic activity of *Phyllanthus niruri* Linn. leaves by homogenous precipitation and turbiditory method. *International Journal of Pharmacy and Pharmaceutical Science*, 6(4), 124–127.
- Kharia, A. A., & Singhai, A. K. (2013). Screening of most effective variables for development of gastroretentive mucoadhesive nanoparticles by Taguchi design. *ISRN Nanomaterials*, 2013, 1–8.
- Kian, V. Y., Mansor, Z., Aziz, A. H. A., Izani, A. S. A., Yew, K. H., & Ahmad, K. (2015). View on traditional complementary medicine of outpatient in Miri hospital. *Sarawak Journal of Pharmacy*, 1(2015), 41–57.
- Kiran, C. R., Rao, D. B., Sirisha, N., & Rao, T. R. (2015). Assessment of phytochemicals and antioxidant activities of raw and germinating *Ceiba pentandra* (kapok) seeds. *Journal of Biomedical Research*, 29(5), 414–419.
- Kishore, D. V., Moosavi, F., & Varma, R. K. (2013). Effect of ethanolic extract of *Portulaca oleracea* Linn. on ethylene glycol and ammonium chloride induced urolithiasis. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(2), 134–140.
- Knoll, T. (2010). Epidemiology, pathogenesis, and pathophysiology of urolithiasis. *European Urology Supplements*, 9(12), 802–806.
- Kozłowska, A., & Szostak-wegierek, D. (2014). Flavonoids - food sources and health benefits. *Roczniki Państwowego Zakładu Higieny*, 65(2), 1–8.
- Krieger, N. S., Asplin, J. R., Frick, K. K., Granja, I., Culbertson, C. D., Ng, A., ... Bushinsky, D. A. (2015). Effect of potassium citrate on calcium phosphate stones in a model of hypercalciuria. *Journal of the American Society of*



Nephrology, 26(12), 3001–3008.

- Kulip, J. (2003). An ethnobotanical survey of medicinal and other useful plants of Muruts in Sabah, Malaysia. *Telopea*, 10(1), 81–98.
- Kulip, J., Fan, L. N., Manshoor, N., Julius, A., Said, I. M., Gisil, J., ... Tukin, W. F. (2010). Medicinal plants in Maliau Basin, Sabah, Malaysia. *Journal of Tropical Biology and Conservation*, 1(6), 21–33.
- Kumar, S. P., Latheef, K. A., & Remashree, A. B. (2014). Ethnobotanical survey of diuretic and antilithiatic medicinal plants used by the traditional practitioners of Palakkad District. *International Journal of Herbal Medicine*, 2(2), 52–56.
- Kumar Shukla, A., Shukla, S., Garg, A., & Garg, S. (2017). A review on anti-urolithiatic activity of herbal folk plants. *Asian Journal of Biomaterial Research*, 3(2), 1–11.
- Kumaran, M. G. S., & Patki, P. S. (2011). Evaluation of an Ayurvedic formulation (Cystone), in urolithiasis: A double blind, placebo-controlled study. *European Journal of Integrative Medicine*, 3(2011), 23–28.
- Kumari, S., & Gupta, A. K. (2018). Antiurolithic activity of *Euphorbia hirta* plant extracts. *International Journal of Pharmacy Research*, 9(2), 23–26.
- Kurian, S., & Josekumar, V. S. (2017). Phytochemical screening, antimicrobial activity and brine shrimp lethality bioassay of different extracts of *Alysicarpus vaginalis* var. *Nummularifolius* (DC.) MIQ.(Family: Fabaceae). *International Journal of Pharmacy and Pharmaceutical Sciences*, 9(1), 1–6.
- Laib, I., & Barkat, M. (2018). Optimization of conditions for extraction of polyphenols and the determination of the impact of cooking on total polyphenolic, antioxidant, and anticholinesterase activities of potato. *Foods*, 7(3), 1–24.
- Lakshmi, N. V. N. (2014). Antilithiatic ethnomedicinal plants used by the native people of Anantapur District, A.P. *International Journal of Research in Applied, Natural and Social Sciences*, 2(7), 61–66.
- Lawal, T. O., Mbanu, A. E., & Adeniyi, B. A. (2014). Inhibitory activities of *Ceiba pentandra* (L.) Gaertn. and *Cordia sebestena* Linn. on selected rapidly growing mycobacteria. *African Journal of Microbiology Research*, 8(24), 2387–2392.
- Leite, A. de S., Islam, M. T., Junior, A. L. G., Sousa, J. M. de C. e, Alencar, M. V. O. B. de, Paz, M. F. C. J., ... Lopes, J. A. D. (2016). Pharmacological properties of cashew (*Anacardium occidentale*). *African Journal of*



Biotechnology, 15(35), 1855–1863.

- Ling, W. T., Liew, F. C., Lim, W. Y., Subramaniam, S., Lynn, B., Direct, F., ... Subramaniam, S. (2018). Shoot induction from axillary shoot tip explants of fig (*Ficus carica*) cv. Japanese BTM 6. *Tropical Life Sciences Research*, 29(2), 165–174.
- Lipkin, M. E., & Preminger, G. M. (2011). Demystifying the medical management of nephrolithiasis. *Reviews in Urology*, 13(1), 34–38.
- Liu, Y., Chen, Y., Liao, B., Luo, D., Wang, K., Li, H., & Zeng, G. (2018). Epidemiology of urolithiasis in Asia. *Asian Journal of Urology*, 5(4), 205–214.
- Lokendrajit, N., Swapana, N., Singh, C. D., & Singh, C. B. (2011). Herbal folk medicines used for urinary and calculi/stone cases complaints in Manipur. *NeBIO*, 2(3), 1–5.
- Madike, L. N., Takaidza, S., & Pillay, M. (2017). Preliminary phytochemical screening of crude extracts from the leaves, stems, and roots of *Tulbaghia violacea*. *International Journal of Pharmacognosy and Phytochemical Research*, 9(10), 1300–1308.
- Mala, A., & Tulika, T. (2015). Therapeutic efficacy of *Centella asiatica* (L.) and *Momordica charantia*: as traditional medicinal plant. *Journal of Plant Science*, 3(1–1), 1–9.
- Malik, N. S. A., & Bradford, J. M. (2008). Recovery and stability of oleuropein and other phenolic compounds during extraction and processing of olive (*Olea europaea* L.) leaves. *Journal of Food, Agriculture and Environment*, 6(2), 8–13.
- Mamillapalli, V., Latha Khantamneni, P., Mohammad, Z., Mathangi, A., Nandigam, N., Namburi, S. M., & Katta, V. (2016). Phytochemical & in vitro antiurolithiatic studies on the leaf extracts of *Bauhinia variegata* Linn. *International Journal of Pharmaceutical Sciences and Research*, 7(10), 4074–4084.
- Manissorn, J., Fong-Ngern, K., Peerapen, P., & Thongboonkerd, V. (2017). Systematic evaluation for effects of urine pH on calcium oxalate crystallization, crystal-cell adhesion and internalization into renal tubular cells. *Scientific Reports*, 7(1), 1–11.
- Maskam, N. A., Mohamad Ravi, N. H., Mohd. Hassan, H. H., & Mohamad Nor, M. (2017). Phytochemicals screening and antioxidant activity of three different

- solvent extracts of *Euodia redleyi* leaves. *Modern Agricultural Science and Technology*, 3(3), 18–21.
- Matsuura, H. N., & Fett-neto, A. G. (2015). Plant alkaloids: main features, toxicity, and mechanisms of action. *Plant Toxins*, (January 2015), 1–15.
- Mawa, S., Husain, K., & Jantan, I. (2013). *Ficus carica* L. (Moraceae): Phytochemistry, traditional uses and biological activities. *Evidence-Based Complementary and Alternative Medicine*, 2013, 1–8.
- Mediani, A., Abas, F., Tan, C. P., & Khatib, A. (2014). Effects of different drying methods and storage time on free radical scavenging activity and total phenolic content of *Cosmos caudatus*. *Antioxidants*, 3(2), 358–370.
- Melo Cavalcante, A. A., Rubensam, G., Picada, J. N., Gomes da Silva, E., Fonseca Moreira, J. C., & Henriques, J. A. P. (2003). Mutagenicity, antioxidant potential, and antimutagenic activity against hydrogen peroxide of cashew (*Anacardium occidentale*) apple juice and cajuina. *Environmental and Molecular Mutagenesis*, 41, 360–369.
- Mikawlawng, K., Kumar, S., & Vandana. (2014). Current scenario of urolithiasis and the use of medicinal plants as antiurolithiatic agents in Manipur (North East India): a review. *International Journal of Herbal Medicine*, 2(1), 1–12.
- Mitra, S. K., Gopumadhavan, S., Venkataranganna, M. V., & Sundaram, R. (1998). Effect of cystone, a herbal formulation, on glycolic acid-induced urolithiasis in rats. *Phytotherapy Research*, 12, 372–374.
- Mittal, A., Tandon, S., Singla, S. K., & Tandon, C. (2015). In vitro studies reveal antiurolithic effect of *Terminalia arjuna* using quantitative morphological information from computerized microscopy. *International Brazilian Journal of Urology*, 41(5), 935–944.
- Moe, O. W., Pearle, M. S., & Sakhaee, K. (2011). Pharmacotherapy of urolithiasis: evidence from clinical trials. *Kidney International*, 79(4), 385–392.
- Mohan, A., Sagar, S., Priya, B., & Bhagyashri, T. (2013). Phytochemical screening, flavonoid content and antioxidant activity of ethanolic extract of *Ceiba pentandra*. *International Research Journal of Pharmacy and Pharmacology*, 4(2), 108–110.
- Monika, J., Anil, B., Aakanksha, B., & Priyanka, P. (2012). Isolation, characterization and in vitro antiurolithiatic activity of cerpegin alkaloid from *Ceropegia bulbosa* var. *lushii* root. *International Journal of Drug Development*



and Research, 4(4), 154–160.

- Mumtaz, F., Raza, S. M., Ahmad, Z., Iftikhar, A., & Hussain, M. (2014). Qualitative phytochemical analysis of some selected medicinal plants occurring in local area of Faisalabad, Pakistan. *Journal of Pharmacy and Alternative Medicine*, 3(3), 5–10.
- Muryanto, S., Hadi, S. D., Purwaningtyas E.F., & Bayuseno A.P. (2014). Effect of *Orthosiphon aristatus* leaves extract on the crystallization behavior of struvite ($\text{MgNH}_4\text{PO}_4 \cdot 6\text{H}_2\text{O}$). In *3 Rd International Conference on Advanced Materials and Practical Nanotechnology (ICAMPN)*.
- Myers, R. H., & Montgomery, D. C. (1995). *Response surface methodology: Process and product optimization using designed experiments*. New York, NY: John Wiley & Sons. Inc.
- Nagal, A., & Singla, R. K. (2013). Herbal resources with antiurolithiatic effects : A review. *Indo Global Journal of Pharmaceutical Sciences*, 3(1), 6–14.
- Nagamani, J. E., Vidya, S. D., & Banu, S. H. (2014). A study on antioxidant and antimicrobial properties of *Bombax ceiba pentandra* seed extract. *World Journal of Pharmacy and Pharmaceutical Sciences*, 3(12), 692–706.
- Nedamani, E. R., Mahoonak, A. S., Ghorbani, M., & Kashaninejad, M. (2014). Antioxidant properties of individual vs. combined extracts of rosemary leaves and oak fruit. *Journal of Agricultural Science and Technology*, 16, 1575–1586.
- Niharika, M., Harshitha, V., Ashwini, P., Srivinya, B., Himabindhu, J., & Ramanjaneyulu, K. (2018). Evaluation of in vitro antiurolithiatic activity of *Chloris barbata*. *International Journal of Current Pharmaceutical Research*, 10(3), 65–67.
- Nik Wil, N. N. A., Noor Adila, M. O., Ibrahim, N. A., & Tajuddin, S. N. (2014). In vitro antioxidant activity and phytochemical screening of *Aquilaria malaccensis* leaf extracts. *Journal of Chemical and Pharmaceutical Research*, 6(12), 688–693.
- Ningombam, D. S., Devi, S. P., Singh, P. K., Pinokiyo, A., & Thongam, B. (2014). Documentation and assessment on knowledge of ethno Medicinal practitioners: A case study on local Meetei healers of Manipur. *IOSR Journal of Pharmacy and Biological Sciences*, 9(1), 53–70.
- Nkouam, G. B., Adjoh, G., Leudeu, C. B. T., Kouebou, C., Tchiegang, C., & Kapseu, C. (2017). Local uses of kapok (*Ceiba pentandra* Gaertn.) tree from



- the Northern Part of Cameroon. *International Journal of Environment, Agriculture and Biotechnology*, 2(4), 2214–2219.
- Nouri, A. I., & Hassali, M. A. (2018). Assessment of the prevalence of kidney stone disease in a Malaysian teaching hospital. *African Journal of Urology*, 24(3), 180–185.
- Nyeem, M. A. Bin, Haque, M. S., Akramuzzaman, M., Siddika, R., Sultana, S., & Islam, B. R. (2017). *Euphorbia hirta* Linn. a wonderful miracle plant of Mediterranean region: A review. *Journal of Medicinal Plants Studies*, 5(3), 170–175.
- Onuh, J., Idoko, G., Yusufu, P., & Onuh, F. (2017). Comparative studies of the phytochemical, antioxidant and antimicrobial properties of cashew leaf, bark and fruits extracts. *American Journal of Food and Nutrition*, 5(4), 115–120.
- O'Connor, S. E. (2010). Alkaloids. In L. Mander & H.-W. Liu (Eds.), *Comprehensive Natural Products II Chemistry and Biology* (Volume 1, pp. 997–1007).
- Olowa, L. F., & Nuñez, O. M. (2013). Brine shrimp lethality assay of the ethanolic extracts of three selected species of medicinal plants from Iligan City, Philippines. *International Research Journal of Biological Sciences*, 2(11), 74–77.
- Ong, H. C., & Norzalina, J. (1999). Malay herbal medicine in Gemencheh, Negri Sembilan, Malaysia. *Fitoterapia*, 70(1), 10–14.
- Pagarra, H., Rahman, R. A., Muis, A., & Arifin, N. (2019). Screening of factors influencing pectin extraction of pomelo peels using 2-level factorial design. *IOP Conference Series: Materials Science and Engineering*, 551, 012127.
- Panche, A., Diwan, A. D., & Chandra, S. R. (2016). Flavonoids: an overview. *Journal of Nutritional Science*, 5(47), 1–15.
- Pandey, A., & Tripathi, S. (2014). Concept of standardization, extraction and pre phytochemical screening strategies for herbal drug. *Journal of Pharmacognosy and Phytochemistry*, 2(5), 115–119.
- Panigrahi, P. N., Dey, S., & Jena, S. C. (2016). Urolithiasis: Critical analysis of mechanism of renal stone formation and use of medicinal plants as antiurolithiatic agents. *Asian Journal of Animal and Veterinary Advances*, 11(1), 9–16.
- Papoutsis, K., Pristijono, P., John, B., Stathopoulos, C. E., Michael, C., Scarlett, C.



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PEPUSIAKAN TUNKU TUNJALINAH

- J., & Vuong, Q. V. (2018). Screening the effect of four ultrasound-assisted extraction parameters on hesperidin and phenolic acid content of aqueous *Citrus pomace* extracts. *Food Bioscience*, 21, 20–26.
- Patel, M. A., Patel, P. K., & Seth, A. K. (2011). Inhibition of calcium oxalate crystallization by the fruit extract of *Piper longum* L. *Pharmacologyonline*, 2, 1169–1177.
- Patel, P.K., Patel, M. A., Saralai, G. M., & T.R. Gandhi. (2012). Antiuro lithiatic effects of *Solanum xanthocarpum* fruit extract on ethylene-glycol induced nephrolithiasis in rats. *Journal of Young Pharmacist*, 4(3), 164–170.
- Patel, Paras K, Patel, M. A., Vyas, B. A., Shah, D. R., & Gandhi, T. R. (2012). Antiuro lithiatic activity of saponin rich fraction from the fruits of *Solanum xanthocarpum* Schrad. & Wendl. (Solanaceae) against ethylene glycol induced urolithiasis in rats. *Journal of Ethnopharmacology*, 144(1), 160–170.
- Pauzi, A. N., Muhammad, N., Sairi, N. H., Tuan Putra, T. N. M., Gul, M. T., Rahim, N. F. A., ... Abdullah, N. (2019). The effect of different solvent extraction towards antiuro lithiatic properties of *Euphorbia hirta* and *Orthosiphon stamineus*. *IOP Conference Series: Earth and Environmental Science*, 269(1), 1–7.
- Pham, H., Nguyen, V., Vuong, Q., Bowyer, M., & Scarlett, C. (2015). Effect of extraction solvents and drying methods on the physicochemical and antioxidant properties of *Helicteres hirsuta* Lour. leaves. *Technologies*, 3(4), 285–301.
- Phatak, R. S., & Hendre, A. S. (2015). In-vitro antiuro lithiatic activity of *Kalanchoe pinnata* extract. *International Journal of Pharmacognosy and Phytochemical Research*, 7(2), 275–279.
- Pizzi, A. (2019). Tannins: perspectives and actual industrial applications. *Biomolecules*, 9(8), 344.
- Prasad, K. V. S. R. G., Bharathi, K., & Srinivasan, K. K. (1993). Evaluation of *Musa paradisiaca* Linn, cultivar - “Puttubale” stem juice for antilithiatic activity in albino rats. *Indian Journal of Physiology and Pharmacology*, 37(4), 337–341.
- Qiu, S. R., Wierzbicki, A., Orme, C. A., Cody, A. M., Hoyer, J. R., Nancollas, G. H., ... De Yoreo, J. J. (2004). Molecular modulation of calcium oxalate crystallization by osteopontin and citrate. *Proceedings of the National Academy of Sciences of the United States of America*, 101(7), 1811–1815.
- Rahim, N. F. A., Muhammad, N., Abdullah, N., & Talip, B. A. (2018). Synergistic



effect of polyherbal formulations on DPPH radical scavenging activity. *Journal of Science and Technology*, 10(2), 116–121.

Rahman, S. F. S. A., Sijam, K., & Omar, D. (2016). *Piper sarmentosum* Roxb.: a mini review of ethnobotany, phytochemistry and pharmacology. *Journal of Analytical & Pharmaceutical Research*, 2(5), 8–10.

Rahmatullah, M., Mukti, I. J., Haque, A. K. M. F., Mollik, M. A. H., Parvin, K., Jahan, R., ... Rahman, T. (2009). An ethnobotanical survey and pharmacological evaluation of medicinal plants used by the Garo Tribal community living in Netrakona district, Bangladesh. *Advances in Natural and Applied Sciences*, 3(3), 402–418.

Raissi, S., & Farsani, R. E. (2009). Statistical process optimization Through multi-response surface methodology. *World Academy of Science, Engineering and Technology*, 39, 280–284.

Rajeh, M. A. B., Zuraini, Z., Sasidharan, S., Latha, L. Y., & Amutha, S. (2010). Assessment of *Euphorbia hirta* L. leaf, flower, stem and root extracts for their antibacterial and antifungal activity and brine shrimp lethality. *Molecules*, 15(9), 6008–6018.

Ramachandran, S., Vamsikrishna, M., Gowthami, K. V., Heera, B., & Dhanaraju, M. D. (2011). Assessment of cytotoxic activity of *Agave cantula* using brine shrimp (*Artemia salina*) lethality bioassay. *Asian Journal of Scientific Research*, 4(1), 90–94.

Ramli, M. R., Milow, P., & Chooi, O. H. (2015). Traditional knowledge of a practitioner in medicinal plants of Masjid Ijok village, Perak, Malaysia. *Studies on Ethno-Medicine*, 9(1), 59–66.

Ramos, G. Q., Da Fonseca De Albuquerque, M. D., Ferreira, J. L. P., Cotta, E. A., & Da Fonseca Filho, H. D. (2016). Wettability and morphology of the leaf surface in cashew tree from the Amazon, Northern Brazil. *Acta Scientiarum - Biological Sciences*, 38(2), 215–220.

Ranganath, M. S., & Vipin, H. (2014). Optimization of process parameters in turning operation using response surface methodology: a review. *International Journal of Emerging Technology and Advanced Engineering*, 4(10), 351–360.

Rashid, A., Khan, A. A., Dar, S., Nabi, N., & Teli, A. R. (2017). Phytochemical and therapeutic properties of *Ficus carica* Linn.: An overview. *International Journal of Pharmaceutical Science and Research*, 2(6), 16–23.



- Rashidi, W. N. A. S. W. M., Muhammad, N., Sairi, N. H., Rahim, N. F. A., Talip, B. A., Abdullah, N., & Bakar, M. F. A. (2019). Phytochemical analysis and in-vitro antiurolithiatic properties of selected Malaysian herbs. *Journal of Advanced Research in Fluid Mechanics and Thermal Sciences*, 1(1), 152–159.
- Rathod, V. D., Fitwe, P., Sarnaik, D., & Kshirsagar, S. N. (2013). In-vitro antiurolithiatic activity of corn silk of *Zea mays*. *International Journal of Pharmaceutical Sciences Review and Research*, 21(2), 16–19.
- Ratkalkar, V. N., & Kleinman, J. G. (2011). Mechanisms of stone formation. *Clinical Reviews in Bone and Mineral Metabolism*, 9(3–4), 187–197.
- Razak, R. N. H. A., Rahman, S. A., Hamdan, A. H., Ramli, R., Muhammad Lokman Md Isad, Muhammad, H., & Hassan, N. F. N. (2019). Evaluation of acute and sub-acute oral toxicity of the aqueous extract of *Aquilaria malaccensis* leaves in Sprague Dawley rats. *Asia Pacific Journal of Molecular Biology and Biotechnology*, 27(1), 20–32.
- Razak, S. A., & Haron, N. W. (2015). Phytosociology of *Aquilaria malaccensis* lamk. and its communities from a tropical forest reserve in Peninsular Malaysia. *Pakistan Journal of Botany*, 47(6), 2143–2150.
- Rewatkar, K., Shende, D. Z., & Wasewar, K. L. (2018). Reactive crystallization of calcium oxalate: population balance modeling. *Chemical and Biochemical Engineering Quarterly*, 32(1), 11–18.
- Ribeiro, D. B. C., Fabricante, J. R., Francisco, S., Referência, C. De, Degradadas, D. Á., & De, P. (2014). Bioinvasion of *Tradescantia zebrina* Heynh. (Commelinaceae) in uplands, State of Paraíba, Brazil. *Brazilian Journal of Biological Sciences*, 1(1), 1–10.
- Roghini, R., & Vijayalakshmi, K. (2018). Phytochemical screening, quantitative analysis of flavonoids and minerals in ethanolic extract of *Citrus paradisi*. *International Journal of Pharmaceutical Sciences and Research*, 9(11), 4859–4864.
- Romero, V., Akpınar, H., & Assimios, D. G. (2010). Kidney stones: a global picture of prevalence, incidence, and associated risk factors. *Reviews in Urology*, 12(2–3), 86–96.
- Roshanak, S., Rahimmalek, M., & Goli, S. A. H. (2016). Evaluation of seven different drying treatments in respect to total flavonoid, phenolic, vitamin C content, chlorophyll, antioxidant activity and color of green tea (*Camellia*



- sinensis* or *C. assamica*) leaves. *Journal of Food Science and Technology*, 53(1), 721–729.
- Saha, S., Shrivastav, P. S., Verma, R. J., Saha, S., Shrivastav, P. S., & Verma, R. J. (2014). Antioxidative mechanism involved in the preventive efficacy of *Bergenia ciliata* rhizomes against experimental nephrolithiasis in rats. *Pharmaceutical Biology*, 52(6), 712–722.
- Saha, S., & Verma, R. J. (2013). Inhibition of calcium oxalate crystallisation in-vitro by an extract of *Bergenia ciliata*. *Arab Journal of Urology*, 11, 187–192.
- Said, O., Khalil, K., Fulder, S., & Azaizeh, H. (2002). Ethnopharmacological survey of medicinal herbs in Israel, the Golan Heights and the West Bank region. *Journal of Ethnopharmacology*, 83(3), 251–265.
- Sairi, N. H., Muhammad, N., Pauzi, A. N., Bakar, M. F. A., Talip, B. H. A., Abdullah, N., ... Ibrahim, N. (2019). The evaluation of antiurolithiatic properties of *Ananas nanus*, *Aquilaria malaccensis*, *Curcuma xanthorrhiza*, *Pandanus atrocarpus* and *Garcinia mangostana*. *Materials Today: Proceedings*, 19, 1145–1150.
- Salminen, A., Lehtonen, M., Suuronen, T., Kaarniranta, K., & Huuskonen, J. (2008). Terpenoids: Natural inhibitors of NFκB signaling with anti-inflammatory and anticancer potential. *Cellular and Molecular Life Sciences Review*, 65, 2979–2999.
- Sanaye, M. M., & Zehra, S. (2018). In-vitro evaluation of anti-urolithiatic activity of leaves of *Alstonia scholaris*. *International Journal of Current Research*, 10(7), 71869–71874.
- Sarah, Q. S., Anny, F. C., & Misbahuddin, M. (2017). Brine shrimp lethality assay. *Bangladesh Journal of Pharmacology*, 12, 186–189.
- Saranya, R., & Geetha, N. (2014). Inhibition of calcium oxalate (CaOx) crystallization in-vitro by the extract of beet root (*Beta vulgaris* L.). *International Journal of Pharmacy and Pharmaceutical Sciences*, 6, 361–365.
- Saranya, S., Nair, A. V., Prathapan, M. P., Neethu, A. S., & Kumar, N. S. (2017). Phytochemical analysis of *Centella asiatica* L. leaf extracts. *International Journal of Advanced Research*, 5(6), 1828–1832.
- Sarigul, N., Korkmaz, F., & Kurultak, İ. (2019). A new artificial urine protocol to better imitate human urine. *Scientific Reports*, 9(1), 1–11.
- Sasidharan, H., Mallya, S. V., Suchitra, P., & Kumar, K. N. S. (2018). In-vitro



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PERPUSTAKAAN FAKULTAS FARMASI

- evaluation of *Scoparia dulcis* Linn. for anti-urolithiatic activity. *The Journal of Phytopharmacology*, 7(3), 284–286.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M., & Latha, L. Y. (2011). Extraction, isolation and characterization of bioactive compounds from plants' extracts. *African Journal of Traditional, Complementary and Alternative Medicines*, 8(1), 1–10.
- Sasikala, V., Radha, S. R., & Vijayakumari, B. (2013). In vitro evaluation of *Rotula aquatica* Lour. for antiurolithiatic activity. *Journal of Pharmacy Research*, 6(3), 378–382.
- Sathish, R., & Jeyabalan, G. (2018). Study of in vitro anti-lithiatic effect of *Ipomoea batatas* (L) leaves and tuberous roots. *Asian Journal of Pharmaceutical and Clinical Research*, 11(2), 427–431.
- Saw, L. G., Chua, L. S. L., Suhaida, M., Yong, W. S. Y., & Hamidah, M. (2010). Conservation of some rare and endangered plants from Peninsular Malaysia. *Kew Bulletin*, 65(4), 681–689.
- Seshagirirao, K., Harikrishnanaik, L., Venumadhav, K., Nanibalu, B., Jamir, K., Ratnamma, B. K., ... Babarao, D. K. (2016). Preparation of herbarium specimen for plant identification and voucher number. *Roxburghia*, 6(1–4), 111–119.
- Seyyedani, A., Yahya, F., Kamarolzaman, M. F. F., Suhaili, Z., Desa, M. N. M., Khairi, H. M., ... Zakaria, Z. A. (2013). Review on the ethnomedicinal, phytochemical and pharmacological properties of *Piper sarmentosum*: scientific justification of its traditional use. *Tang [Humanitas Medicine]*, 3(3), 1–32.
- Shang, H., Zhou, H., Li, R., Duan, M., & Wu, H. (2017). Extraction optimization and influences of drying methods on antioxidant activities of polysaccharide from cup plant (*Silphium perfoliatum* L.). *PloS One*, 12(8), 1–18.
- Sharifa, A. A., Jamaludin, J., Kiong, L. S., Chia, L. A., & Osman, K. (2012). Anti-urolithiatic terpenoid compound from *Plantago major* Linn. (ekor anjing). *Sains Malaysiana*, 41(1), 33–39.
- Sharma, D., Dey, Y. N., Sikarwar, I., Sijoria, R., Wanjari, M. M., & Jadhav, A. D. (2016). In-vitro study of aqueous leaf extract of *Chenopodium album* for inhibition of calcium oxalate and brushite crystallization. *Egyptian Journal of Basic and Applied Sciences*, 3(2), 164–171.



- Sharma, N., Tanwer, B. S., & Vijayvergia, R. (2011). Study of medicinal plants in Aravali regions of Rajasthan for treatment of kidney stone and urinary tract troubles. *International Journal of PharmTech Research*, 3(1), 110–113.
- Shetty, B. S., Udupa, S. L., & Udupa, A. L. (2008). Biochemical analysis of granulation tissue in steroid and *Centella asiatica* (Linn) treated rats. *Pharmacologyonline*, 2, 624–632.
- Shi, J., Yu, J., Pohorly, J., Young, J. C., Bryan, M., & Wu, Y. (2003). Optimization of the extraction of polyphenols from grape seed meal by aqueous ethanol solution. *Journal of Food Agriculture & Environment*, 1(2), 42–47.
- Shivani, M., Spandana, K., Himabindhu, J., & Ramanjaneyulu, K. (2018). Evaluation of in-vitro antiurolithiatic activity of *Piper nigrum*. *International Journal of Pharmaceutical Sciences and Medicine*, 3(8), 43–50.
- Silberstein, J., Lakin, C. M., & Parsons, J. K. (2008). Shock wave lithotripsy and renal hemorrhage. *Rev*, 10(3), 236–241.
- Silva, E. M., Souza, J. N. S., Rogez, H., Rees, J. F., & Larondelle, Y. (2006). Antioxidant activities and polyphenolic contents of fifteen selected plant species from the Amazonian region. *Food Chemistry*, 101(3), 1012–1018.
- Simanjutak, M. (2008). *Ekstraksi and fraksinasi komponen ekstrak daun tumbuhan senduduk (Melsatoma malabathricum) serta pengujian efek sediaan krim terhadap penyembuhan luka bakar*. Indonesia: Universitas Sumatera Utara, Sumatera.
- Singh, A., Prakash, J., Meghwal, P. R., & Ranpise, S. A. (2015). Fig (*Ficus carica* L.). In S. N. Ghosh (Ed.), *Breeding of Underutilized Fruit Crops Part I* (Edition I, pp. 149–179). New Delhi: Jaya Publishing House.
- Singh, S., Gautam, A., Sharma, A., & Batra, A. (2010). *Centella asiatica* (L.) plant with immense medicinal potential but threatened. *International Journal of Pharmaceutical Sciences Review and Research*, 4(2), 9–17.
- Soloman George, D., Razali, Z., & Somansundram, C. (2016). Physiochemical changes during growth and development of pineapple (*Ananas comosus* L. Merr. cv. Sarawak). *Journal of Agricultural Science and Technology*, 18, 491–503.
- Solomon, A., Andemariam, A., Tseghehannes, F., Aron, H., Tesfagaber, M., Tesfay, D., & Kaushik, A. (2019). Antiurolithiatic activity of the leaf extracts of *Maerua angolensis*. *Archives of Pharmacy and Pharmacology Research*, 2(2),



1–6.

- Soundararajan, P., Mahesh, R., Ramesh, T., & Hazeena Begum, V. (2006). Effect of *Aerva lanata* on calcium oxalate urolithiasis in rats. *Indian Journal of Experimental Biology*, 44(December 2006), 981–986.
- Sreenevasan, G. (1990). Urinary stones in Malaysia - its incidence and management. *The Medical Journal of Malaysia*, 45(2), 92–112.
- Sri Widyawati, P., Budianta, T. D. W., Kusuma, F. A., & Wijaya, E. L. (2014). Difference of solvent polarity to phytochemical content and antioxidant activity of *Pluchea indicia* less leaves extracts. *International Journal of Pharmacognosy and Phytochemical Research*, 6(4), 850–855.
- Srikanth, M., Kasala, S., Ramanjaneyulu, K., Himabindhu, J., & Kiran, J. S. (2017). Evaluation of in vitro anti urolithiatic activity of *Acacia farnesiana*. *World Journal of Pharmacy and Pharmaceutical Sciences*, 6(11), 1080–1084.
- Srikaran, R., & Dulanjali, S. S. (2019). Evaluation of in-vitro anti-urolithiatic activity of methanolic extract of *Cucumis melo* seeds on calcium oxalate crystals. *International Journal of Current Pharmaceutical Research*, 11(1), 18–20.
- Srivastava, N., Chauhan, A. S., & Sharma, B. (2012). Isolation and characterization of some phytochemicals from Indian traditional plants. *Biotechnology Research International*, 2012.
- Suarez, M., & Youssef, R. (2015). Potassium Citrate: treatment and prevention of recurrent calcium nephrolithiasis. *Journal of Clinical Nephrology and Research*, 2(1), 1015–1021.
- Sulaiman, S. F., Yusoff, N. A. M., Eldeen, I. M., Seow, E. M., Sajak, A. A. B., Supriatno, & Ooi, K. L. (2011). Correlation between total phenolic and mineral contents with antioxidant activity of eight Malaysian bananas (*Musa* sp.). *Journal of Food Composition and Analysis*, 24(1), 1–10.
- Suleiman, D., Ibrahim, U. I., & Saravanan, D. (2018). In-vitro antilithiasis activity of *Melastoma malabathricum* Linn. *Journal of Natural Sciences Research*, 8(8), 20–26.
- Tabugo, S. R. M., Rampola, R. B., Jr., E. V., & B, M. R. S. (2013). Assessment of the safety of tawa-tawa (*Euphorbia hirta* L.) decoction as alternative folkloric medicine. *Global Journal of Medicinal Plant Research*, 1(2), 177–183.
- Tan, J. B. L., Yap, W. J., Tan, S. Y., Lim, Y. Y., & Lee, S. M. (2014). Antioxidant



- content, antioxidant activity, and antibacterial activity of five plants from the Commelinaceae family. *Antioxidants*, 3, 758–769.
- Tédonga, L., Dzeufieta, P. D. D., Dimoa, T., Asongalem, E. A., Sokengc, S. N., Flejoub, J.-F., ... Kamtchouinga, P. (2007). Acute and subchronic toxicity of *Anacardium occidentale* Linn (Anacardiaceae) leaves hexane extract in mice. *African Journal of Traditional, Complementary and Alternative Medicines*, 4(2), 140–147.
- Telekar, Y. P., Gund, K. A., & Kale, S. D. (2013). Antiuro lithiatic activity of corn silk extract in mice. *International Journal of Universal Pharmacy and Bio Sciences*, 2(4), 65–77.
- Tiselius, H. (2000). Stone incidence and prevention. *Brazilian Journal of Urology*, 26(5), 452–462.
- Tiwari, A., Soni, V., Londhe, V., Bhandarkar, A., Bandawane, D., & Nipate, S. (2012). An overview on potent indigenous herbs for urinary tract infirmity: Urolithiasis. *Asian Journal of Pharmaceutical and Clinical Research*, 5(1), 7–12.
- Touhami, M., Laroubi, A., Elhabazi, K., Loubna, F., Zrara, I., Eljahiri, Y., ... Chait, A. (2007). Lemon juice has protective activity in a rat urolithiasis model. *BMC Urology*, 7(18), 1–10.
- Turjaman, M., Santoso, E., & Sumarna, Y. (2006). Arbuscular Mycorrhizal fungi increased early growth of gaharu wood of *Aquilaria malaccensis* and *A. crasna* under greenhouse conditions. *Indonesian Journal of Forestry Research*, 3(2), 139–148.
- Ueda, H., Kaneda, N., Kawanishi, K., Alves, S. M., & Moriyasu, M. (2002). A new isoflavone glycoside from *Ceiba pentandra* (L.) GAERTNER. *Chemical and Pharmaceutical Bulletin*, 50(3), 403–404.
- Uma, D. B., Ho, C. W., & Wan Aida, W. M. (2010). Optimization of extraction parameters of total phenolic compounds from Henna (*Lawsonia inermis*) leaves. *Sains Malaysiana*, 39(1), 119–128.
- Upadhyay, N., Tiwari, S. K., Seth, A., & Maurya, S. K. (2017). Effect of *Averrhoa carambola* Linn. fruit on ethylene glycol induced urolithiasis in rats. *Current Research Journal of Pharmaceutical and Allied Sciences*, 01(01), 2–8.
- Varicola, K., Metla, S., Syamala, U. S., & Rajulapati, S. (2017). Assessment of potential antiuro lithiatic activity of some selected medicinal plants by in vitro



- techniques. *Acta Scientific Pharmaceutical Sciences*, 1(2), 38–42.
- Velioglu, Y. S., Mazza, G., Gao, L., & Oomah, B. D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *Journal of Agricultural and Food Chemistry*, 46, 4113–4117.
- Velu, V., Fuloria, N., Fuloria, S., Panda, J., Panda, B. P., & Malipeddi, H. (2018). In-vitro and in-vivo anti-urolithiatic activity of terpenoid-rich ethyl acetate extract of rhizomes of *Curcuma zedoaria*. *Studies on Ethno Medicine*, 12(1), 31–39.
- Waghulde, S., Kale, M. K., & Patil, V. (2019). Brine shrimp lethality assay of the aqueous and ethanolic extracts of the selected species of medicinal plants. *Proceedings*, 41, 47.
- Wang, C.-M., Yeh, K.-L., Tsai, S.-J., Jhan, Y.-L., & Chou, C.-H. (2017). Anti-proliferative activity of triterpenoids and sterols isolated from *Alstonia scholaris* against non-small-cell lung carcinoma cells. *Molecules*, 22, 2119.
- Wang, J., Sun, B., Cao, Y., Tian, Y., & Li, X. (2008). Optimisation of ultrasound-assisted extraction of phenolic compounds from wheat bran. *Food Chemistry*, 106(2), 804–810.
- Wang, S., Yu, Z., Wang, C., Wu, C., Guo, P., & Wei, J. (2018). Chemical constituents and pharmacological activity of agarwood and *Aquilaria* plants. *Molecules*, 23(2), 1–21.
- Wang, Y., Fang, X., An, F., Wang, G., & Zhang, X. (2011). Improvement of antibiotic activity of *Xenorhabdus bovienii* by medium optimization using response surface methodology. *Microbial Cell Factories*, 10(1), 98.
- Wattanathorn, J., Wannanon, P., Muchimapura, S., Thukham-mee, W., Tong-un, T., & Polyiam, P. (2019). Toxicity evaluation of *Anacardium occidentale*, the potential aphrodisiac herb. *BioMed Research International*, 2019, 1–20.
- Worcester, E. M., & Coe, F. L. (2008). Nephrolithiasis. *Primary Care*, 35(2), 369–391.
- Wu, D., Liu, W., Han, Q., Wang, P., Xiang, X., Ding, Y., ... Qin, W. (2019). Extraction optimization, structural characterization, and antioxidant activities of polysaccharides from Cassia Seed (*Cassia obtusifolia*). *Molecules*, 24, 1–16.
- Yadav, N., Yadav, R., & Goyal, A. (2014). Chemistry of terpenoids. *International Journal of Pharmacy and Pharmaceutical Sciences Review and Research*, 27(2), 272–278.



- Yadav, R. D., Jain, S. K., Alok, S., Mahor, A., Bharti, J. P., & Jaiswal, M. (2011). Herbal plants used in the treatment of urolithiasis: a review. *International Journal of Pharmaceutical Sciences and Research*, 2(6), 1412–1420.
- Yi, S., Su, Y., Qi, B., Su, Z., & Wan, Y. (2010). Application of response surface methodology and central composite rotatable design in optimizing the preparation conditions of vinyltriethoxysilane modified silicalite/polydimethylsiloxane hybrid pervaporation membranes. *Separation and Purification Technology*, 71(2), 252–262.
- Yuan, X., Zeng, Y., Nie, K., Luo, D., & Wang, Z. (2015). Extraction optimization, characterization and bioactivities of a major polysaccharide from *Sargassum thunbergii*. *PloS One*, 10(12), 1–11.
- Zahid, I. H., Bawazir, A. S., & Naser, R. (2013). Plant based native therapy for the treatment of Kidney stones in Aurangabad (MS). *Journal of Pharmacognosy and Phytochemistry*, 1(6), 189–193.
- Zaidan, M. R. S., Noor Rain, A., Badrul, A. R., Adlin, A., Norazah, A., & Zakiah, I. (2006). In vitro screening of five local medicinal plants for antibacterial activity using disc diffusion method. *Tropical Biomedicine*, 22(2), 167–170.
- Zainol, N. A., Voo, S. C., Sarmidi, M. R., & Aziz, R. A. (2008). Profiling of *Centella asiatica* (L.) urban extract. *The Malaysian Journal of Analytical Sciences*, 12(2), 322–327.
- Zakaria, F., Talip, B. A., Kahar, E. E. M., Muhammad, N., Abdullah, N., & Basri, H. (2020). Solvent used in extraction process of agarwood: A systematic review. *Food Research*, 4(3), 731–737.
- Zakaria, M., & Mohd, M. A. (2015). *Traditional Malay Medicinal Plants*. Kuala Lumpur, Malaysia: Institut Terjemahan & Buku Malaysia Berhad.
- Zakaria, Z. A., Nor, S. R. N. R. M., Kumar, G. H., Ghani, Z. D. F. A., Sulaiman, M. R., Devi, G. R., ... Fatimah, C. A. (2006). Antinociceptive, anti-inflammatory and antipyretic properties of *Melastoma malabathricum* leaves aqueous extract in experimental animals. *Canadian Journal of Physiology and Pharmacology*, 84(12), 1291–1299.
- Zakaria, Z. A., Rofiee, M. S., Mohamed, A. M., Teh, L. K., & Salleh, M. Z. (2011). In vitro antiproliferative and antioxidant activities and total phenolic contents of the extracts of *Melastoma malabathricum* leaves. *Journal of Acupuncture and Meridian Studies*, 4(4), 248–256.



- Zambrano, M. V., Dutta, B., Mercer, D. G., MacLean, H. L., & Touchie, M. F. (2019). Assessment of moisture content measurement methods of dried food products in small-scale operations in developing countries: A review. *Trends in Food Science and Technology*, 88, 484–496.
- Zeng, X., Xi, Y., & Jiang, W. (2019). Protective roles of flavonoids and flavonoid-rich plant extracts against urolithiasis: A review. *Critical Reviews in Food Science and Nutrition*, 59(13), 2125–2135.
- Zwenger, S., & Basu, C. (2008). Plant terpenoids: applications and future potentials. *Biotechnology and Molecular Biology Reviews*, 3(1), 1–7.
- Zzaman, W. (2016). *Optimization of antioxidant extraction from jackfruit (Artocarpus heterophyllus Lam.) seeds using response surface methodology*. Ghent University.



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PERPUSTAKAAN TUNKU TUN AMINAH